



Cannabidiolic Acid (CBDA) - Properties and Effects

Gerhard Nahler

Zusammenfassung

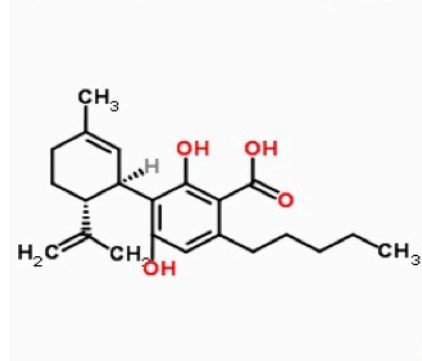
Cannabidiolsäure (kurz CBDA) ist das Hauptcannabinoid in Industriehanf. Unter Temperatur- und Lichteinfluß wandelt sich CBDA in Cannabidiol (CBD) um. Diese Umwandlung ist in Extrakten jedoch keineswegs vollständig; abhängig von den Bedingungen finden sich stets mehr oder weniger hohe Anteile an CBDA neben CBD. Der CBDA-Gehalt im Produkt ist höher, wenn früh geerntet wird und wenn sämtliche Prozesse möglichst wenig Licht- und Temperatureinflüssen ausgesetzt sind.

CBDA ist zwar in seinen Eigenschaften dem CBD ähnlich, hat aber unterschiedliche Wirkungen. Im Gegensatz zu CBD ist CBDA ein Hemmer der Cyclooxygenase 2 (COX-2), also einem Enzym, das für die Bildung von pro-inflammatorischen Metaboliten wie Prostaglandine verantwortlich ist. Auch betreffend einer Interaktion mit dem „Serotonin-Rezeptor“ 5-HT1A lässt CBDA eine wenigstens 10x höhere Wirksamkeit bei Übelkeit, Erbrechen und Angstzuständen erwarten. Schließlich wird vermutet, dass CBDA die eigentlich antimikrobiell-wirksame Substanz in Cannabinoidgemischen ist. Gegen Tumorzellen ist CBDA ebenfalls wirksam, wenn auch, in vitro, schwächer als CBD.

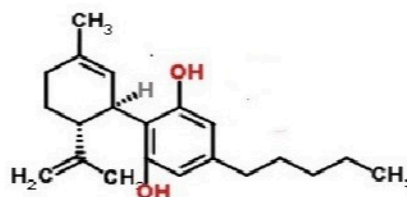
Besonders erwähnenswert ist, dass CBDA die Bioverfügbarkeit von CBD etwa um einen Faktor zwei erhöht. Anders ausgedrückt, ein Extrakt mit einem etwa 1:1 Anteil von CBDA: CBD hat nicht nur die volle Wirkung eines „reinen“ CBD-Extrakts, sondern zusätzlich die Vorteile von CBDA. Tierversuche lassen vermuten, dass CBDA besser bioverfügbar sein dürfte, doch fehlen derzeit Daten am Menschen.

Introduction

Cannabidiolic acid (CBDA) was the earliest discovered cannabinoid acid. It was first isolated in 1955 (Izzo et al., 2009; Brenneisen 2007). In fresh plant material, particularly in unripe samples, 95% of CBD exists as its acid (Turner et al., 1980). In industrial hemp extracts and products that have been prepared by keeping exposure to light and heat to a minimum, CBDA is the main cannabinoid, surpassing concentrations of cannabidiol (CBD) by a factor of 10 and more. But even if heated, the decarboxylation process is not complete.



CBDA

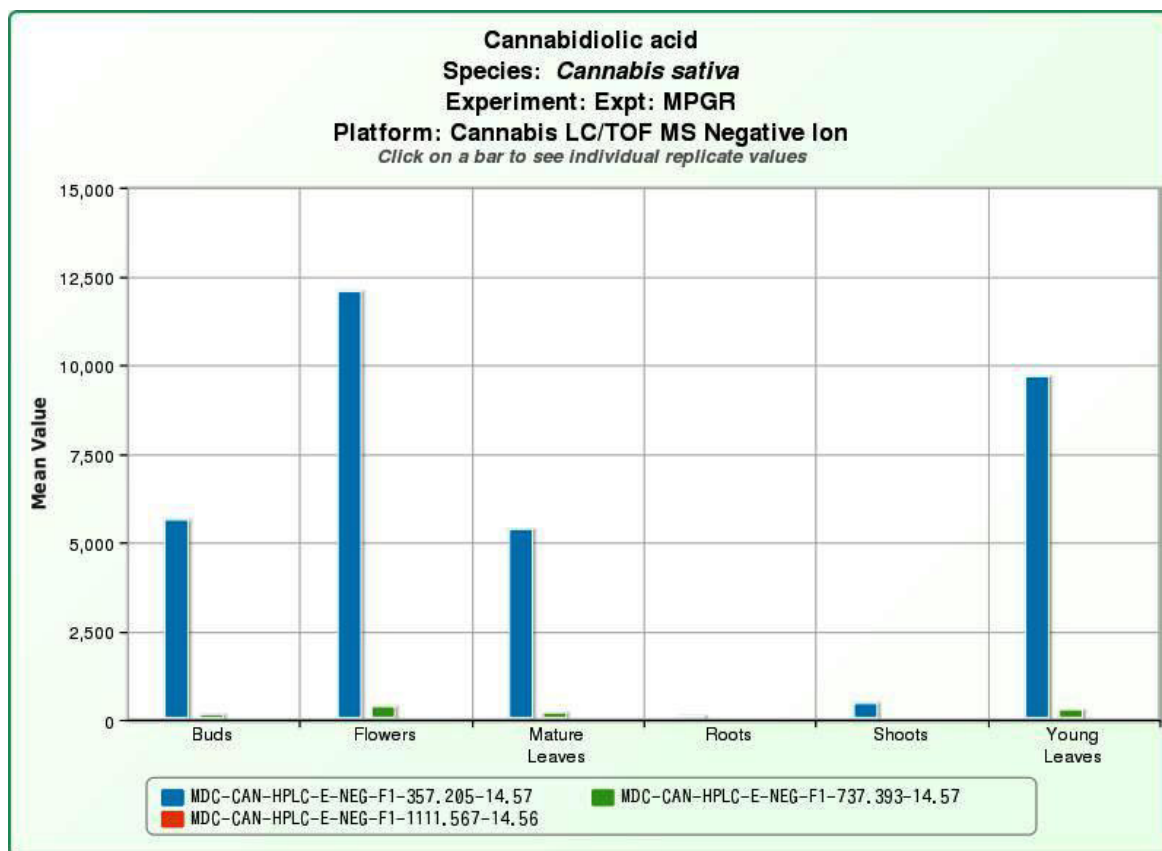


CBD

Solubility CBDA [water, 25°C (est)]: 3.158 ng/ml (<http://www.thegoodscentscompany.com/data/rw1399421.html>)

Solubility CBD [water, 25°C (est)]: 5.509 ng/ml (<http://www.thegoodscentscompany.com/data/rw1399301.html>)

CBDA content is highest in flowers and young leaves:



From: Medicinal Plant Metabolomics Resource, http://metnetweb.gdcb.iastate.edu/mpmr_public/metabolites/?id=88913;

Conversion of CBDA to CBD

The decarboxylation of CBDA to CBD is non-enzymatically; CBD increases slowly after harvesting, during storage or through heating and exposure to light which are the two most important factors. In unheated extracts, concentrations of cannabinoid acids (mcg/ml) can be about 100 times that of the decarboxylated forms:

	CBD	CBDA	CBDA/CBD	THC	THCa	THCa/THC
Extract High in CBD - Unheated	26	3,500	135	48	3,300	69
<i>Heated</i>	<i>1,900</i>	<i>0.9</i>	<i>0.0005</i>	<i>1,100</i>	<i>9.3</i>	<i>0.008</i>
Extract A, High in THC - Unheated	3.9	28	7	90	14,500	161
<i>Heated</i>	<i>29</i>	<i>17</i>	<i>0.586</i>	<i>10,060</i>	<i>150</i>	<i>0.015</i>

Heated for 7 minutes at 200°C; From: Verhoecx et al., 2006; concentrations (mcg/ml);

To note, D-9-tetrahydrocannabinolic acid exists in two forms, THCA-A and THCA-B, but only THCA-A is relevant; (here abbreviated as THCa in order to avoid confusion with the metabolite of THC, 11-nor-delta9-tetrahydrocannabinol-9-carboxylic acid (delta9-THC-COOH) that is sometimes also abbreviated as THCA).

As can be seen from the table above, decarboxylation is only partial, even when heated at 200°C for 7 minutes (Verhoeckx et al., 2006). Therefore, small amounts of CBDA but also of D-9-terahydrocannabinolic acid (THCa) can still be found, even when extracts are heated. The fact that the presence of THCa (THCA-A) can be detected in blood and urine after smoking marijuana has been proposed as a (forensic) possibility to demonstrate exposure to cannabis (Kyriazou et al., 2014).

Very little is known about the speed of this transformation of CBDA to CBD; it seems to be about half the speed of transformation of THCa. Therefore, it is estimated to be in the order of 6 months (dark, room temperature) to several years (dark, +4°C; Wang et al., 2016). Light has the greatest influence. After heating at 60°C for two hours (pH = 7) no inactivation of CBDA has been observed whereas heating at 100°C reduced the activity by 50% (Gal et al., 1969).

Pharmacokinetics

CBDA shows a good bioavailability as determined in male Wistar rats. Following to single administration, absolute bioavailability of i.p. injected pure CBDA was 80% with 10 mg/kg, 62% with 1 mg/kg and 36% with 0.1 mg/kg; oral bioavailability was 19% after 10 mg/kg. After repeated oral administration, bioavailability of CBDA increased to 39% after 10mg/kg, and was 7% after i.p. administration of 0.1mg/kg (Jones et al., 2014). Oral bioavailability of CBDA-botanical drug substance (extract) was slightly higher than of pure CBDA which points toward enhancement of absorption by natural byproducts.

At present, no human data with pure CBDA are available. Intriguingly, the bioavailability of CBD was roughly twice as high when about 42% of the total amount of CBD (decarboxylated CBD + CBDA) of an extract administered orally, was in form of CBDA (Eichler et al., 2012). Therefore, not only the bioavailable amount of CBD of such a partially decarboxylated extract is equivalent to a fully decarboxylated extract, but offers the additional advantages of functional CBDA. At present, it is not clear whether CBDA enhances the absorption of CBD or reduces its first pass effect in the liver.

Targets and effects of CBDA

CBDA exerts pharmacological properties by its own and interacts with a number of targets of the endocannabinoid system. CBDA has weak activity on CB1 and CB2 receptors, is a partial agonist of the Transient Receptor Potential type TRPA1, of TRPV1, an antagonist of TRPM8 and a cyclooxygenase (COX-2) inhibitor (IC50: 2.2 mM; Izzo et al., 2009). CBDA binds strongly to Hydroxy-Tryptamin receptor 5-HT1A and weakly to 5-HT5A; the corresponding botanical drug substance (BDS) does not interact (Jones et al., 2014). In contrast to THC and CBD, CBDA inhibits cyclooxygenases (COX-2 about 10-times stronger than COX-1; Takeda et al., 2008; Ruhaak et al., 2011). Cyclooxygenases are enzymes that produce powerful inflammatory mediators such as prostaglandins. The effect on COX-2 is about 10-times weaker than diclofenac ("Voltaren™"), but the amount that can be taken safely is more than 10-times higher. From that, anti-inflammatory effects and a possible use for pain control might be delineated. More details are given below:

Target	Concentration	Role of CBDA	Reference
COX-1	IC50: 20-470 mcM	Weak Inhibitor; Cyclooxygenase has a pro-inflammatory role	Ruhaak et al., 2011; Takeda et al., 2008;
COX-2	IC50: 2.2mcM	Inhibitor; COX-2 has a pro-inflammatory role	Takeda et al., 2008;
DAGL	IC50: 19.4mcM	Weak Inhibitor;	De Petrocellis 2011;
GPR55	1-10 mcM	Involved in inflammation and tumour development; (CBDA slightly better antagonist than CBD, EC50 between 1 and 3 mcM)	Anavi-Göffer et al., 2012; Izzo 2009
5-HT1A ("serotonine receptor")	< 1mcM	Involved in anxiolysis, attenuation of nausea, vomiting, cerebral infarction; CBDA is at least 10-times more potent than CBD; (CBD ≈80% displacement at 16 mcM)	Bolognini et al., 2013; Cascio et al., 2014 ; Izzo 2009
5-HT5A	15mcM	Weak inhibitor (IC50 15mcM; Ki 750 nM);	Bolognini et al., 2013
TRPA1	1-10 mcM; EC50: 12 mcM	Agonist, attenuates nociception, (CBD, EC50, 0.096 mcM; more effective),	De Petrocellis et al., 2011, 2008; Izzo 2009
TRPM8	1-10 mcM	Antagonist (EC50: 0.9-1.9 mcM); (CBD, IC50: 0.08-0.14-0.503 mcM; more effective)	De Petrocellis et al., 2008, 2011; Izzo 2009
TRPV1 ("Capsaicin receptor")	> 10mcM; EC50: 10 mcM	Agonist, marginally weaker than CBD; activation induces apoptosis in cancer cells and evokes vasodilation; (CBD, EC50: 1-3 mcM)	De Petrocellis et al., 2011; Ligresti et al., 2006; Izzo 2009
TRPV4	1-10 mcM	mainly involved in sensory (nociceptive) functions; similar effective as CBD;	De Petrocellis et al., 2012

DAGL - diacylglycerol lipase (DAGL α & DAGL β); produce 2-AG from the hydrolysis of diacyl-glycerols;

GPR55 – G-Protein coupled Receptor 55;

5-HT - Hydroxy-Tryptamin receptor

TRP – Transient Receptor Potential, A-ankrin type, M-melastatin type, V-vanilloid type;

Properties and potential uses of CBDA

Condition	Reference
Anxiolytic, in vivo, 0.05-5 mg/kg; <i>p.o.</i>	Brierley et al.2016;
Anti-inflammatory (Inhibition of COX-2); analgesic	Ruhaak et al., 2011; Takeda et al., 2008
inhibits gastrointestinal contractions & transit <i>in vivo</i> ,	Cluny et al., 2011
reduced nausea and vomiting <i>in vivo</i> (Lithium-, cisplatin-induced) in dosages around 0.1 – 0.5mg/kg	Bolognini et al., 2011; Rock et al., 2016
Antitumor effects, <i>in vitro</i> & <i>in vivo</i> (breast-, gastric-, glioma-, liver-, pancreatic-, renal-, thyroid cancer cell lines);	Ligresti et al., 2006; Takeda et al., 2016, 2015, 2012; Hill et al., 2014
Antimicrobial activity <i>in vitro</i> (Gram-positive bacteria including methicillin-, tetracycline-, macrolide-resistant Staphylococcus aureus strains *)	Grlic 1962; Farkas, Andrassy 1976;

*) From: https://pubchem.ncbi.nlm.nih.gov/compound/Cannabidiolic_acid#section=Top

In various animal models, already low doses of CBDA prevented vomiting (shrews, 0.1 or 0.5 mg/kg i.p.) and nausea (rats, 0.01 or 0.1 mg/kg i.p.) with greater potency than CBD (Bolognini

et al., 2013), enhanced suppression of Lithium (Li-)-induced nausea in comedication with ineffective doses of metoclopramide (0.1 µg CBDA/kg; Rock, Parker 2013) and suppressed anticipatory nausea (Rock et al., 2014). Combination with THC or odansetron has synergistic effects (Rock, Parker 2015; Rock et al., 2015).

CBDA and CBD have inhibitory actions on the intestines of *Suncus murinus* that are not neuronally mediated or mediated via CB(1) or CB(2) receptors (Cluny et al., 2011). CBDA exhibits antitumor effects on various cancer cell types and can inhibit the migration of highly aggressive breast cancer (Takeda et al., 2017; Takeda et al., 2016; Takeda et al., 2012). It was however less potent (in vitro) than CBD (Ligresti et al., 2006; Hill et al., 2014).

CBDA was found to inhibit Gram-positive bacteria and to have a high inhibiting effect on the spores of *Bacillus cereus*. It seems to represent the main antibiotic agent in cannabis resin. Its sporostatic effect is roughly equivalent to that of the antibiotics nisin and tylosin (Grlic 1962; Farkas, Andrassy 1976) and has been proposed as food preservative (Gal et al., 1969).

Toxicity of CBDA

Genotoxicity of pure CBDA and its corresponding botanical drug substance (BDS) were tested in vitro (bacterial reverse mutation assay, Ames test) and in vivo (rat micronucleus test, RMT) up to the maximum doses. There was no evidence of genotoxicity neither of pure CBDA nor for its corresponding BDS. The only observations were transient salivation (pure CBDA) or ataxia and decreased activity (CBDA-BDS) at the highest dose. Signs returned to normal by the 3rd day (Ayerkawa et al., 2014).

No other data are currently available.

Literature:

- Anavi-Goffer S**, Baillie G, Irving AJ, et al: Modulation of L-alpha-lysophosphatidylinositol / GPR55 mitogen-activated protein kinase (MAPK) signaling by cannabinoids. *J Biol Chemistry* 2012;287:91-104.
- Ayerkawa L**, Stott C, Duncan M, et al: Evaluation of the genotoxicity of cannabidiolic acid. 24th Annual Symposium of the International Cannabinoid Research Society, Baveno, Italy, June 28 – July 3, 2014. Available at <http://www.icrs.co/SYMPOSIUM.2014/ICRS2014.PROGRAMME.pdf>. Accessed on August 29, 2016.
- Bolognini D**, Rock EM, Cluny NL, et al: Cannabidiolic acid prevents vomiting in *Suncus murinus* and nausea-induced behaviour in rats by enhancing 5-HT1A receptor activation. *Br J Pharmacol* 2013 Mar;168(6):1456-70.
- Brenneisen R**: Chemistry and analysis of phytocannabinoids and other cannabis constituents. Chap.2, in: Marijuana and the Cannabinoids, Mahmoud A. ElSohly, ed., Humana Press, Totowa, New Jersey 2007.
<http://books.google.at/books?hl=de&lr=&id=G_XtSXd4JH0C&oi=fnd&pg=PA17&ots=MHsxcMRi6J&sig=mf6EPTcJObfWpgVRn7gsh-kBtvo#v=onepage&q&f=false>
- Brierley D**, Samuels, Duncan M, et al.: Neuromotor tolerability and behavioural characterisation of cannabidiolic acid, a phytocannabinoid with therapeutic potential for anticipatory nausea. *Psychopharmacology* 2016;233(2):243-254.
- Cascio MG**, Zamberletti E, Marini P, et al: The phytocannabinoid, D9-tetrahydrocannabivarin, can act through 5-HT1A receptors to produce anti-psychotic effects. 24th Annual Symposium of the International Cannabinoid Research Society, Baveno, Italy, June 28 – July 3, 2014. Available at <http://www.icrs.co/SYMPOSIUM.2014/ICRS2014.PROGRAMME.pdf>. Accessed on August 29, 2016.
- Cluny NL**, Naylor RJ, Whittle BA, Javid FA: The effects of cannabidiolic acid and cannabidiol on contractility of the gastrointestinal tract of *Suncus murinus*. *Archives of Pharmacol Research* 2011 September, 34(9):1509-1517.
- De Petrocellis L**, Ligresti A, Moriello AS, et al: Effects of cannabinoids and cannabinoid-enriched Cannabis extracts on TRP channels and endocannabinoid metabolic enzymes. *Br J Pharmacol* 2011 Aug;163(7):1479-1494.
- Eichler M**, Spinedi L, Unfer-Grauwiler S, et al: Heat exposure of Cannabis sativa extracts affects the pharmacokinetic and metabolic profile in healthy male subjects. *Planta Med* 2012 May;78(7):686-691.
- Farkas J**, Andrassy E: The sporostatic effect of cannabidiolic acid. *Acta Alimentaria Academiae Scientiarum Hungaricae*. 1976;5(1):57-67.
- Gál IE**, Vajda O, Bekes I: Prüfung einiger Eigenschaften der Cannabidiolsäure mit besonderer Berücksichtigung ihrer Anwendbarkeit bei der Lebensmittelkonservierung. *Nahrung* 1969;13(6):515-522.

- Grlic L:** A comparative study on some chemical and biological characteristics of various samples of cannabis resin. UNODC - Bulletin on Narcotics – 1962, Issue 3 – 004; Pages: 37 to 46. Available at http://www.unodc.org/unodc/en/data-and-analysis/bulletin/bulletin_1962-01-01_3_page005.html. Accessed on Aug 30, 2016.
- Grunefeld et al.** Psychopharmacological Activity of Some Substances Extracted from Cannabis sativa. *Electroencephalography and Clinical Neurophys* 27(2);219-220. 1969.
- Hill TDM, Stott CG, Duncan M:** An in vitro evaluation of combinations of phytocannabinoids in pancreatic, gastric, renal, bladder and liver cancer cell lines. 24th Annual Symposium of the International Cannabinoid Research Society, Baveno, Italy, June 28 – July 3, 2014. Available at <http://www.icrs.co/SYMPOSIUM.2014/ICRS2014.PROGRAMME.pdf>. Accessed on August 29, 2016.
- Izzo AA, Borrelli F, Capasso R, DiMarzo V, Mechoulam R:** Non-psychotropic plant cannabinoids: new therapeutic opportunities from an ancient herb. *Trends in Pharmacological Sciences* 2009;30(10): 515-527.
- Jones N, Ayerakawa L, Stott C, Duncan M:** Pharmacological properties, pharmacokinetics and bioavailability of pure cannabidiolic acid and cannabidiolic botanical drug substance. 24th Annual Symposium of the International Cannabinoid Research Society, Baveno, Italy, June 28 - July 3, 2014. Available at <http://www.icrs.co/SYMPOSIUM.2014/ICRS2014.PROGRAMME.pdf>. Accessed on August 29, 2016.
- Jung, J. et al.** Detection of Delta9-tetrahydrocannabinolic acid A in human urine and blood serum by LC-MS/MS. *J Mass Spectrom* 2007; Mar; 42(3): 354-60.
- Kyriazou A, Chatzinikolaou F, Koutsoukis D, et al.:** Δ9-tetrahydrocannabinolic acid A: a reliable marker for differentiating between the consumption of illegal cannabis products and legal, medical Δ9-THC. *Aristotle University Medical Journal* 2014;41(2):20-21.
- Ligresti A, Moriello AS, Starowicz K, Matias I, Pisanti S, De Petrocellis L, Laezza C, Portella G, Bifulco M, Di Marzo V:** Antitumor activity of plant cannabinoids with emphasis on the effect of cannabidiol on human breast carcinoma. *J Pharmacol Exp Ther* 2006 Sep;318(3):1375-1387.
- Rock EM, Parker LA:** Cannabinoids as potential treatment for chemotherapy-induced nausea and vomiting. *Frontiers in Pharmacology* 2016 July;7, Article 221, 10 pages.
- Rock EM, Limebeer CL, Parker LA:** Effect of combined doses of D9-tetrahydrocannabinol (THC) and cannabidiolic acid (CBDA) on acute and anticipatory nausea using rat (Sprague-Dawley) models of conditioned gaping. *Psychopharmacol. (Berlin)* 2015 Dec;232(24):4445-4454.
- Rock EM, Parker LA:** Synergy between cannabidiol, cannabidiolic acid, and Δ⁹-tetrahydrocannabinol in the regulation of emesis in the *Suncus murinus* (house musk shrew). *Behav Neurosci* 2015 Jun;129(3):368-370.
- Rock EM, Limebeer CL, Navaratnam R, et al:** A comparison of cannabidiolic acid with other treatments for anticipatory nausea using a rat model of contextually elicited conditioned gaping. *Psychopharmacol (Berl)* 2014 Aug;231(16):3207-15.
- Rock EM, Parker LA:** Suppression of lithium chloride-induced conditioned gaping (a model of nausea-induced behaviour) in rats (using the taste reactivity test) with metoclopramide is enhanced by cannabidiolic acid. *Pharmacol Biochem Behav.* 2013 Oct;111:84-89.
- Ruhaak LR, Felth, J., Karlsson, P.C., Rafter, J.J., Verpoorte, R., Bohlin L:** Evaluation of the cyclooxygenase inhibiting effects of six major cannabinoids isolated from Cannabis sativa. *Biol. Pharm. Bull.* 2011; 34:774–778.
- Takeda S, Himeno T, Kakizoe K, et al:** Cannabidiolic acid-mediated selective down-regulation of c-fos in highly aggressive breast cancer MDA-MB-231 cells: possible involvement of its down-regulation in the abrogation of aggressiveness. *J Nat Med.* 2017 Jan; 71(1):286-291.
- Takeda S, Himeno T, Kakizoe K, et al:** Cannabidiolic acid-mediated selective down-regulation of c-fos in highly aggressive breast cancer MDA-MB-231 cells: possible involvement of its down-regulation in the abrogation of aggressiveness. *J Natural Medicines* 2016;pp 1-6, doi:10.1007/s11418-016-1030-0.
- Takeda S, Okazaki H, Kohro-Ikeda E, et al :** DNA microarray analysis of genes in highly metastatic 4T1E/M3 murine breast cancer cells following exposure to cannabidiolic acid. *Fundamental Toxicological Sciences* 2015;2(2):89-94.
- Takeda S, Okajima S, Miyoshi H, et al:** Cannabidiolic acid, a major cannabinoid in fiber-type cannabis, is an inhibitor of MDA-MB-231 breast cancer cell migration. *Toxicology Letters* 2012 Nov 15;214(3):314-319
- Takeda S, Misawa K, Yamamoto I, Kazuhito WatanabeK:** Cannabidiolic Acid as a Selective Cyclooxygenase-2 Inhibitory Component in Cannabis. *Drug Metabolism and Disposition* 2008;36(9):1917-1921.
- Turner CE, Elsohly Ma, Boeren EG:** Constituents of Cannabis sativa L. XVII. A review of the natural constituents. *J Natural Prod* 1980 Mar-Apr; 43(2): 169-234.
- Verhoeckx KCM, Korthout HAAJ, Ehlert KA et al.:** Unheated Cannabis sativa extracts and its major compound THC-acid have potential immuno-modulating properties not mediated by CB1 and CB2 receptor coupled pathways. *Int Immunopharmacol* 2006 Apr;6(4):656-665.
- Wang M, Wang YH, ElSohly A, et al.:** Decarboxylation Study of Acidic Cannabinoids: A Novel Approach Using Ultra-High-Performance Supercritical Fluid Chromatography/Photodiode Array-Mass Spectrometry. *Cannabis and Cannabinoid Research* 2016; Vol. 1.1, 2016; 10 pages. DOI: 10.1089/can.2016.0020.